# **GlnaFit** A user guide

March 2015

An illustration of how to model bacterial inactivation curves using GInaFit v1.6, a freeware add-in for Microsoft<sup>®</sup> Excel designed for bridging predictive modelling approaches and end-users in the food industry or research groups.

Further documentation can be found in Geeraerd *et al.* (2005).

GInaFit can be downloaded from http://cit.kuleuven.be/biotec/ginafit.php



Prepared by Z. van der Waal

## CONTENTS

1. Produce the survival curve2	2
2. Determine the shape of the bacterial inactivation curve2	2
3. Select candidate model(s)	4
4. Run and assess with GInaFiT	
Goodness of fit6	6
	6
	6
5. Interpretation and inferences	
Shape of the curve and associated model type6	6
Shoulder effect6	5
	6
	6
6. Further reading 8	8

Figure 1: determine the shape of the bacterial inactivation curves using key descriptive features \_\_\_\_\_\_3

Table 1: Candidate models per type of inactivation shape	4
Table 2: Elements for interpretation of parameters involved in GlnaFit models	7

**1. Produce the survival curve** that relates the log-concentration of heat-injured cells ( $log_{10}$  of N, expressed in CFU.mL<sup>-1</sup>) with duration of the treatment (time in minutes).

**2. Determine the shape of the bacterial inactivation curve** using key descriptive features using the diagram on Figure 1, a flow chart of the procedure to assess the shape of the survival curve. The description starts with the decline phase: is the decline phase linear? If so, does the curve have a shoulder; if not, does it have a tail? The survival curve will be categorised as one of the ten possible shapes described and numbered by Geeraerd *et al.* (2005).

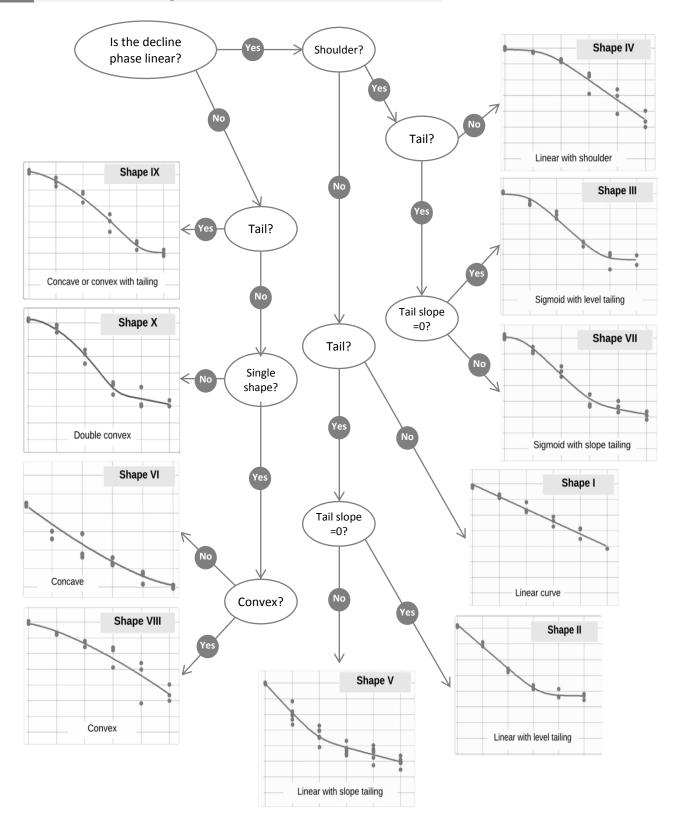


Figure 1: determine the shape of the bacterial inactivation curves using key descriptive features.

The curve can take one of ten shapes: linear (Shape I), linear with tailing (Shape II), sigmoidal-like (Shape III), linear with a preceding shoulder (Shape IV), biphasic (Shape V), concave (Shape VI), convex (Shape VII), convex or concave with a tail (Shape IX), double convex or concave (Shape X).

Inactivation curves used for illustration are triplicates of repeated measurements of the concentration (logCFU.ml<sup>-1</sup>) of heat-injured cells of various strains of *Campylobacter* over time (minutes).

Table 1: Candidate models per type of inactivation shape.

Shape	Applicable models in GInafit	GInafit menu item	Implications/assumptions from model structure	Cited literature in GInafit
Shape I	Log-linear regression	#1	<ul> <li>Traditional first-order inactivation kinetics equation</li> <li>Assumes all cells have equal sensitivity and inactivation depends on random chance of receiving lethal treatment</li> </ul>	Bigelow and Esty, 1920
Linear	Models by Geeraerd <i>et al.,</i> 2000	#2	Note: these models can produce survival curves that replicate the classical Log-linear regression depending on	Albert & Mafart, 2005 Anonymous, 2000 Cerf, 1977
	Weibull + tail Biphasic model	#3	the underlying response, but may not be the most appropriate choice for modelling the data	Cole <i>et al.,</i> 1993 Geeraerd <i>et al.,</i> 2000
Shape II Biphasic, level tailing	Log-linear + tail	#2	Traditional first-order inactivation kinetics equation with added tailing parameter	Geeraerd <i>et al.,</i> 2000
	Biphasic model	#4	Assumes one initially major subpopulation, that is more sensitive to stress (initial decline), and one minor subpopulation that is more resistant to stress (tail)	Cerf, 1977
Shape III Sigmoidal, level tailing	Log-linear + shoulder + tail	#2	Traditional first-order inactivation kinetics equation with added parameters for tailing and shoulder	Geeraerd <i>et al.,</i> 2000 Greenacre <i>et al.,</i> 2003 Marquenie <i>et al.,</i> 2003 Mossel <i>et al.,</i> 1995
Shape IV Linear, shoulder	Log-linear + shoulder	#2	Traditional first-order inactivation kinetics equation with added shoulder parameter	Geeraerd <i>et al.,</i> 2000 Mossel <i>et al.,</i> 1995

**3. Select candidate model(s)**: Identify one or several model types relevant to the shape of the survival curve and available in GInaFit.

Shape	Applicable models in GInafit	GInafit menu item	Implications/assumptions from model structure	Cited literature in GInafit
Shape V Biphasic, slope tailing	Biphasic model	#4	Assumes one initially major subpopulation, that is more sensitive to stress (initial steep constant decline), and one minor subpopulation that is more resistant to stress (final, smoother constant decline)	Cerf, 1977
Shape VI Concave	Weibull	#3	<ul> <li>Shape parameter (<i>p</i>&lt;1) describes the concave shape</li> <li>Non-autonomous model, <i>i.e.</i> D varies with time</li> </ul>	Mafart <i>et al.,</i> 2002 Peleg & Cole, 1998 Van Boekel, 2002
Shape VII Sigmoidal, slope tailing	Biphasic + shoulder	#4	<ul> <li>Most complex shape</li> <li>Combines biphasic model and Geeraerd <i>et al.</i> (2000) shoulder parameter</li> </ul>	Geeraerd <i>et al.,</i> 2005 Whiting, 1993
Shape VIII Convex	Weibull	#3	<ul> <li>Shape parameter (<i>p</i>&lt;1) describes the convex shape</li> <li>Non-autonomous model: <i>D</i> varies with time</li> </ul>	Mafart <i>et al.,</i> 2002 Peleg & Cole, 1998 Van Boekel, 2002
Shape IX Convex or concave, tailing	Weibull + tail	#3	<ul> <li>Shape parameter describes the convex/concave shape</li> <li>Non-autonomous model, <i>i.e.</i> D varies with time</li> </ul>	Albert & Mafart, 2005 Albert & Mafart, 2003
Shape X Double convex	Mixed Weibull	#3	Assumes one initially major subpopulation, that is more sensitive to stress (first wave), and minor subpopulation	Coroller <i>et al.,</i> 2006

that is more resistant to stress (second wave)

**4.** Run and assess with GlnaFiT: See Geeraerd *et al.* (2012) manual for general installation and use. Develop relevant model(s); the software will test for mis-uses (*e.g.* when applying a model with tailing on data not having a tail, or when the number of parameters is too high in comparison with the number of data points). Model outputs include indicators of:

Goodness of fit:	Precision:	Variation explained:
The Root mean sum of squared error (RMSE) quantifies the goodness of fit for both linear and non linear models via the difference between prediceted and observed values. Best fit indicated when this value is close to zero.	<ul> <li>calculate SD, the standard deviation of data.</li> <li>RMSE &gt;&gt; SD: model not capturing trend, too flexible (overfitting, following noise)</li> <li>RMSE &lt;&lt; SD: model not capturing trend, not flexible enough (underfitting, not following signal)</li> </ul>	R-square adjusted coefficient of determination is the proportion of variation in the inactivation curve that is explained by the model, with penalization for irrelevant parameters (overfitting). Best fit indicated when this value is close to 1.

5. Interpretation and inferences: alongside numeric model outputs, inferences can be drawn using:

**Shape of the curve and associated model type** may vary within and between bacterial strains, be influenced by stress intensity (frequently concave may become convex or sigmoidal), physiological state of cells, phase of growth (exponential or stationary phase), pre-stress conditions *etc.* In addition to the elements for interpretation in Table *pp*4-5, also consider:

**Shoulder effect** suggests initial resistance to stress (see Albert & Mafart, 2005

**Tailing effect** can suggest varying levels of resistance, for instance due to mixed populations, clumping, protective effect of the suspension medium

**Parameter estimates** can provide insights into destructive mechanisms. resistance to stress, residual cell concentration, treatment efficiency *etc*. Literature provides various illustration and theoretical background for model parameters (*e.g.* Coroller *et al.*, 2006; Xion *et al.*, 1999; Buchanan *et al.*, 1993 on  $t_{4D}$ ); elements for interpretation are summarised in Table 2 for the model parameters in GInaFit.

#### Model Focus for interpretation Parameter Log-linear with shoulder Duration of Time before decrease: initial SI Log-linear with shoulder and tail shoulder effect resistance to stress Biphasic with shoulder and tail Log-linear with shoulder Speed of decrease per time unit (constant, after shoulder and/or Log-linear with tail Log-linear with shoulder and tail First order before tail) $K_{max1}$ and $K_{max2}$ for speed of decrease k<sub>max</sub> inactivation rate for initially major and minor constant Biphasic with tail Biphasic with shoulder and tail populations (constant, after shoulder and/or before tail) Log-linear with shoulder Log-linear with tail Log-linear with shoulder and tail Initial inoculum concentration Initial inoculum Weibull $N_0$ (similar to population size in concentration Weibull with tail constant volume) Mixed Weibull Biphasic with tail Biphasic with shoulder and tail Log-linear with tail Residual cell concentration after Starting point of $N_{\text{res}}$ Log-linear with shoulder and tail stabilisation at the end of the tail Weibull with tail decrease Weibull Treatment lethality, close to the Weibull with tail classical D-value Time to first log- $\delta_1$ and $\delta_2$ for first and second δ reduction of first subpopulation; subpopulation<sub>1</sub> is Mixed Weibull subpopulation more sensitive to stress than subpopulation<sub>2</sub> when $\delta_1 < \delta_2$ Weibull Shape of Weibull with tail *p*<1 for convex, *p*>1 for concave р inactivation curve **Mixed Weibull** $\alpha$ is defined as the logit of f and is equivalent to $\alpha = \log_{10}(N_{01}/N_{02})$ , and Fraction of first subpopulation the $\alpha$ value then is close to the Mixed Weibull α remaining in total graphic difference between $\log_{10}(N_0)$ population and the logarithm of the population size where the inflection is observe Fraction of initially Biphasic with tail Major subpopulation is the least f major Biphasic with shoulder and tail resistant of both subpopulation Treatment lethality: time needed for a 4log reduction of N<sub>0</sub>. Automatically reported for data sets logcycles of While relevance of the classical Dcovering at least 4 decimal $t_{4D}$ reduction value is restricted to logreductions linear curves; t<sub>4D</sub> applies to log-linear

# Table 2: Elements for interpretation of parameters involved in GlnaFit models.

and non-log linear survival curves.

### 6. Further Reading

Albert I. & P. Mafart 2005. A modified Weibull model for bacterial inactivation, International Journal of Food Microbiology 100: 197-211.

Albert I. & P. Mafart 2003. A modified Weibull model for bacterial inactivation. Van Impe J.F.M., A.H. Geeraerd, I. Leguérinel & P. Mafart (Eds.), Predictive Modelling in Foods - Conference Proceedings, 90-5682-400-7, Katholieke Universiteit Leuven/BioTeC, Belgium: 143-145.

Bigelow W.D. & J.R. Esty 1920. The thermal death point in relation to typical thermophylic organisms, Journal of Infectious Diseases 27: 602-617.

Buchanan R.L., Golden M.H. & Whiting R.C. 1993. Differentiation of the effects of pH and lactic or acetic acid concentration on the kinetics of Listeria monocytogenes inactivation, Journal of Food Protection 56: 474-478.

Cerf O. 1977. A review. Tailing of survival curves of bacterial spores, Journal of Applied Microbiology 42: 1-19.

Cole M.B., K.W. Davies, G. Munro, C.D. Holyoak & D.C. Kilsby, 1993. A vitalistic model to describe the thermal inactivation of *Listeria monocytogenes*. Journal of Industrial Microbiology 12: 232–239.

Coroller L., I. Leguérinel, E. Mettler, N. Savy & P. Mafart, 2006. General model, based on two mixed Weibull distributions of bacterial resistance, for describing various shapes of inactivation curves. Applied and Environmental Microbiology 72: 6493-6502.

Geeraerd A.H., 2013. GInaFiT: Geeraerd and Van Impe Inactivation model Fitting Tool. Version 1.6 – Office 2010 & 2007, March 2012.

Geeraerd A.H., V.P. Valdramidis & J.F. Van Impe, 2005. GInaFiT, a freeware tool to assess non-log-linear microbial survivor curves. International Journal of Food Microbiology 102: 95-105.

Geeraerd A.H., C.H. Herremans & J.F. Van Impe 2000. Structural model requirements to describe microbial inactivation during a mild heat treatment, International Journal of Food Microbiology 59: 185-209.

Greenacre E.J., T.F. Brocklehurst, C.R. Waspe, D.R. Wilson & P.D.G. Wilson. 2003. *Salmonella enterica* serovar Typhimurium and *Listeria monocytogenes* Acid Tolerance Response induced by organic acids at 20 °C: optimization and modelling. Applied and Environmental Microbiology 69 (7): 3945–3951. Mafart P., I. Leguérinel, O. Couvert & L. Coroller, 2010. Quantification of spore resistance for assessment and optimization of heating processes: a never-ending story. Food Microbiology: 568-572.

Mafart P., O. Couvert, S. Gaillard & I. Leguerinel 2002. On calculating sterility in thermal preservation methods: application of the Weibull frequency distribution model, International Journal of Food Microbiology 72: 107-113.

Marquenie D., A.H. Geeraerd, J. Lammertyn, C. Soontjens, J.F. Van Impe, C.W. Michiels & B.M. Nicolaï. 2003. Combinations of pulsed white light and UV-C or mild heat treatment to inactivate conidia of *Botrytis cinerea* and *Monilinia fructigena*. International Journal of Food Microbiology 85: 185–196.

Mossel D.A.A., J.E.L. Corry, C.B. Struijk & R.M. Baird, 1995. Essentials of the Microbiology of Foods. Wiley, West Sussex, UK.

Peleg M. & M.B. Cole, 1998. Reinterpretation of microbial survival curves. Critical Reviews in Food Science and Nutrition 38: 353–380.

Pruitt K.M. & D.N. Kamau, 1993. Mathematical models of bacterial growth, inhibition and death under combine stress conditions. Journal of Industrial Microbiology 12: 221-231.

Sakkaf A.A., G. Jones & J. Mawson, 2010. Survival of New Zealand relevant *Campylobacter* strains. Final Report: N0174/06, Prepared for NZFSA, March 2010.

Sichel C., J. Blanco, S. Malato & P. Fernandez-Ibanez, 2007. Effects of experimental conditions on *E. coli* survival during solar photocatalytic water disinfection. Journal of Photochemistry and photobiology A: Chemistry, 189(2): 239-246.

Teixeira A.A. & A. Rodriguez, 1990. Microbial population dynamics in bioprocess sterilization. Enzyme and Microbial Technology 12: 469-473.

Van Boekel M.A.J.S. 2002. On the use of the Weibull model to describe thermal inactivation of microbial vegetative cells. International Journal of Food Microbiology 74: 139–159.

Whiting R.C. 1993. Modeling bacterial survival in unfavorable environments. Journal of Industrial Microbiology 12: 240–246.

Xiong R., G. Xie, A. E. Edmondson & M. A. Sheard, 1999. A mathematical model for bacterial inactivation. International journal of food microbiology 46: 45-55

8